## 139. <sup>13</sup>C-NMR Spectra of Cannabinoids

Part 2<sup>1</sup>)

#### Side-Chain Substituted Tetrahydrocannabinols and Synthetic Intermediates

by Ingo Franke, Burkhard Schmidt, Wolfgang Dietrich<sup>a</sup>) and Michael Binder<sup>\*2</sup>)

Institute für Physiologische Chemie und <sup>a</sup>) Analytische Chemie (SC-Abteilung) der Ruhr-Universität Bochum, Postfach 102148, D-4630 Bochum 1

(19.XII.83)

### Summary

The <sup>13</sup>C-NMR spectra of eight semi-synthetic side-chain substituted cannabinoids and of eight synthetic intermediates were analyzed in detail. Assignments of the signals are based on their chemical shifts, splitting patterns in <sup>1</sup>H-off-resonance decoupling experiments, incremental calculations, and model considerations.

**Introduction.** – We have reported the analysis of the <sup>13</sup>C-NMR spectra of several semi-synthetic cannabinoids substituted in the dibenzopyrane moiety [1]. These compounds were grouped around  $\Delta^{9(11)}$ -tetrahydrocannabinol (=  $\Delta^{9(11)}$ -THC), a cannabinoid model compound devoid of psychotropic activity in the rhesus monkey [2]. The major objective of our investigations being the elucidation of the molecular mechanism of action of the psychotropically active cannabinoid  $\Delta^9$ -THC,  $\Delta^{9(11)}$ -THC was employed to differentiate non-specific, lipophilicity-mediated effects which  $\Delta^{9}$ - and  $\Delta^{9(11)}$ -THC share from specific, receptor-mediated effects which should be unique to  $\Delta^9$ -THC [3]. The next step was the synthesis of 17,18-didehydro- $\Delta^8$ -THC 8 via 18-bromo- $\Delta^8$ -THC 7 as precursor for  $[17,18-{}^{3}H_{2}]-\Delta^{8}$ -THC in order to conduct binding studies to the putative THC receptor. As tools for the characterization of this receptor, cannabinoid affinity gels were developed employing 17-methyl- $\Delta^{8}$ -THC-18-oic acid 2 as affinity ligand, and the synthesis [4] and some biological properties [5] of the corresponding N-ethyl amide 6 which represents the closest approximation to the final affinity unit were reported. The Scheme summarizes the structural formulas of the intermediates and cannabinoids involved in the syntheses of 17,18-didehydro- $\Delta^8$ -THC 8 and the amides 5 and 6 (cf. [4]) for which we now report also the <sup>13</sup>C-NMR data.

The assignments of the <sup>13</sup>C-NMR signals are based on general <sup>13</sup>C-NMR shift theory [6], <sup>1</sup>H-off-resonance decoupling, and incremental calculations [7].

<sup>&</sup>lt;sup>1</sup>) Part 1: see [1].

<sup>&</sup>lt;sup>2</sup>) Deceased on February 15<sup>th</sup>, 1984.









9 R = H **10**  $R = CH_3$ 

11 R = H12 R = CH<sub>3</sub>

13



14b (1*E*)

# Analysis of the <sup>13</sup>C-NMR Spectra.

Experimental. The measurements were performed at 62.9 MHz on a Bruker-WM-250 NMR spectrometer by use of proton-noise and off-resonance decoupling. All spectra were recorded in CDCl<sub>3</sub> solution at 300 K with more than 240 ppm sweep width.

Tat	ole 1. 13 C-NMR Shifts	(in ppm Downfield	from Me <sub>4</sub> Si) and	Splitting Pattern of	<sup>c</sup> Cannabinoid Com	<i>spounds.</i> Predicted	shifts in parenthe	ses.
C	1	2	3	4	5	6	7	80
- 1	155.0 s	155.9	155.2	155.1	155.9	155.8	154.9	154.8
2	107.7 d	107.6	107.6	107.6	107.8	107.8	107.7	107.6
3	141.6 s	141.6	141.5	141.7	141.4	141.6	142.1	142.2
4	109.9 d	109.8	109.6	109.9	109.0	109.4	110.1	110.1
4a	154.8 5	154.8	154.7	154.9	154.6	154.8	154.9	154.7
6	76.7 s	76.6	76.6	76.6	76.5	76.6	76.7	76.7
6a	44.9 d	44.9	44.8	44.9	45.0	45.1	44.9	44.8
7	27.9 1	27.9	27.8	27.9	27.9	28.0	27.8	27.9
8	119.3 d	119.3	119.2	119.3	119.1	119.2	119.3	119.3
6	134.7 s	134.7	134.7	134.7	134.9	135.0	134.7	134.7
10	36.0 t	36.0	35.8	36.1	36.4	36.1	36.1	36.0
10a	31.6 <i>d</i>	31.6	31.5	31.6	31.6	31.8	31.6	31.5
10b	110.9 s	110.9	110.8	110.8	110.9	110.9	110.8	110.6
11	23.4 q	23.3	23.4	23.3	23.3	23.4	23.4	23.5
12	27.4 q	27.5	27.5	27.5	27.5	27.6	27.5	27.5
13	18.4 <i>q</i>	18.4	18.4	18.4	18.4	18.5	18.5	18.5
14	34.9 1 (35.0)	35.1 t (35.5)	34.9 1 (35.0)	35.2 t (35.5)	35.0 1 (34.6)	35.4 t (35.1)	35.1 ( (35.5)	34.8 ( (35.3)
15	30.1 t (28.3)	28.2 t (27.9)	30.2 1 (28.3)	28.4 t (27.9)	30.2 ( (27.9)	28.5 t (27.5)	29.9 t (30.0)	30.0 t (31.1)
16	24.3 t (24.2)	33.1 t (31.2)	24.5 t (24.2)	33.4 t (31.2)	25.3 t (24.8)	33.9 t (31.8)	27.8 t (27.9)	33.3 t (32.2)
17	33.9 1 (33.3)	39.1 d (39.0)	35.9 1 (35.8)	39.3 d (41.5)	35.9 t (35.2)	41.5 d (40.9)	33.5 t (33.6)	138.6 d (138.1)
18	179.8 s	182.1 s	174.5 s	177.4 s	173.7 s	176.9 s	32.7 t (32.4)	114.6 t (114.4)
CH <sub>3</sub> -C(17)		16.8 q (15.0)		$17.0 \ q$		17.9 q (15.0)		
CH <sub>3</sub> 0			51.5 q	51.4 q				
NCH <sub>2</sub> CH <sub>3</sub>					34.4 t	34.4 1		
NCH, CH,					14.6 <i>q</i>	14.8 <i>q</i>		

Helvetica Chimica Acta - Vol. 67 (1984)

	Table 2. 13 C-NMR Shifts	(in ppm Downfield	from Me <sub>4</sub> Si) and	Splitting Pattern o,	f Synthetic Interme	ediates. Predicted s	shifts in parenthese	ss.
С	6	10	11	12	13	I 4a	14b	15
1	173.0 \$	176.2 s	175.6 s	178.6 s	35.5 t (35.5)	132.0 <i>d</i>	130.5 d	36.1 t (35.5)
2	33.9 t (33.9)	39.6 d (41.7)	35.1 t (36.0)	39.4 d (41.5)	29.9 1 (30.0)	129.6 d	130.1 d	28.8 / (30.7)
3	23.9 t (25.3)	32.3 t (34.4)	24.3 t (24.1)	33.0 t (31.2)	27.7 t (27.9)	25.1 t (28.7)	28.7 t (28.7)	25.5 t (25.5)
4	129.9 d	129.6 d	30.1 t (29.8)	28.2 t (28.6)	33.7 t (33.6)	29.4 t (31.0)	29.2 t (31.0)	30.8 t (32.7)
5	130.7 d	130.7 d	33.8 t (35.0)	35.3 t (35.5)	32.6 / (32.4)	67.0 t (67.1)	66.8 t (67.1)	67.3 t (62.5)
ľ'	139.0 s <sup>a</sup> )(138.1)	139.2 s <sup>a</sup> ) (138.1)	145.0 s (145.0)	144.9 s (145.0)	145.7 s (145.0)	139.2 s (137.9)	139.6 s (137.9)	144.7 s (144.5)
2',6'	108.0 d (104.7)	105.1 d (104.7)	108.0 d (108.0)	107.9 d (108.0)	108.3 d (108.0)	106.7 d (103.9)	103.9 d (103.9)	106.5 d (105.5)
3',5'	159.7 s (157.0)	159.8 s (157.0)	156.6 s (156.0)	156.7 s (156.0)	156.4 s (156.0)	160.4 s (157.8)	160.7 s (157.8)	160.4 s (159.4)
4,	100.8 d (95.2)	101.0 d (95.2)	105.5 d(100.2)	100.6 d (100.2)	100.5 d(100.2)	98.7 d (95.6)	99.2 d (95.6)	97.5 d (96.3)
1"	$136.9 \ s^{a})(141.5)$	137.0 s <sup>a</sup> ) (141.5)				158.9 s (158.7)	158.9 s (158.7)	158.8 s (158.7)
2",6"	127.2 d(127.1)	127.4 d (127.1)				114.4 d (113.8)	$114.4 \ d(113.8)$	114.2 d(113.8)
3",5"	128.4 d(128.5)	128.5 d (128.5)				129.3 d (129.4)	129.3 d (129.4)	129.5 d (129.4)
4"	127.7 d (127.3)	127.8 d (127.3)				120.4 d (120.4)	120.4 d (120.4)	120.1 d (120.4)
$CH_{3}-C(2)$		16.5 q (15.3)		16.7 q (16.1)				
OCH <sub>3</sub>	51.3 q	51.4 q	51.9 q	51.8 q		55.0 q	55.0 q	
OCH2C6H5	(0.69) 1 6.69	70.1 t (69.0)						
<sup>a</sup> ) Assignme	nts based on integral area	of the resp. signals						

1236

Discussion. The <sup>13</sup>C-NMR data of compounds **1–8** (all with (6aR, 10aR)-configuration; **2**, **4**, and **6**: 17*RS*) are summarized in *Table 1*. All eight molecules share an identical tetrahydrodibenzopyrane moiety, and the signals of the respective <sup>13</sup>C-atoms appear within  $\pm 0.5$  ppm at the same position as those of the parent compound  $A^{8}$ -THC [1]. The chemical shifts of the side chain C-atoms (C(14) to C(18)) were estimated using the *Grant-Paul* relation [6] considering compounds **1**, **3**, and **5** as butane, **2**, **4**, **6**, and **7** as pentane, and **8** as propane derivatives.

A set of increments (Z) for the complete tetrahydrodibenzopyrane moiety was obtained by comparing the <sup>13</sup>C-NMR chemical shifts of pentane and  $\Delta^8$ -THC:  $Z_{\alpha} = 22.0$ ,  $Z_{\beta} = 8.3$ ,  $Z_{\gamma} = -2.6$ , and  $Z_{\delta} = 0.2$  ppm. Employing the reported Z-values for -COOH, -COOR, -CONR<sub>2</sub>, -HC=CH<sub>2</sub>, -Br, and the necessary steric corrections, the predicted  $\delta$ -values (see *Table 1*) suffice to assign the observed signals to the corresponding C-atoms. All assignments are confirmed by the off-resonance splitting patterns of the resp. signals.

The <sup>13</sup>C-NMR data of the synthetic intermediates 9–15 are given in *Table 2*. The observed chemical shifts of compounds 11-13 and 15 agree well with the values obtained by incremental calculations. For the spectra of the olefinic compounds 9, 10, 14a, and 14b, the calculations prove sufficient to locate all signals arising from aliphatic and aromatic C-atoms, but fail with the olefinic C-atoms, the predicted values differing as much as 6 ppm from the observed ones. Since of all four compounds the <sup>1</sup>H-NMR signals of the respective olefinic protons are less than 0.5 ppm apart (cf. [4]), selective frequency decoupling could not be employed. Therefore, the assignments for the olefinic C-atoms are based on theoretical considerations: according to the <sup>1</sup>H-NMR spectra [4], the olefinic double bond of both compounds 9 and 10 is in trans-configuration. Except for the  $CH_1$ -group at C(2), 9 and 10 are identical. Thus, the <sup>13</sup>C-NMR chemical shift increment induced by the introduction of this CH<sub>3</sub>-group when going from 9 and 10 should influence the position of the signal of the  $\gamma$ -carbon atom C(4) but not the one arising from the  $\delta$ -carbon atom C(5). Therefore, the signals at 130.7 ppm in 9 and at 130.7 ppm in 10 are assigned to C(5) and the signals at 129.9 ppm (9) and 129.6 ppm (10) to C(4). For compound 14, spectra of both the cis-(14a) and *trans*-form (14b) were recorded. According to the *Dreiding* models of 14a and 14b, C(2) of 14a is twisted out of the plane of the aromatic ring by at least 50°. Compared to 14b, this places C(2) and especially C(3) of 14a into the aromatic shielding cone, shifting the signals of these C-atoms 0.5 and 3.6 ppm upfield. Since conjugation is interrupted, C(1) and C(2')/C(6') are deshielded by 1.5 and 2.8 ppm, respectively.

#### REFERENCES

- [1] M. Binder, I. Franke, B. Schmidt & W. Dietrich, Helv. Chim. Acta 65, 807 (1982).
- [2] M. Binder, H. Edery & G. Porath, 'Marihuana: Biological Effects', eds. G. Nahas and W.D.M. Paton, Pergamon Press, Oxford, 1979, p.71.
- [3] M. Binder, F.-J. Witteler, B. Schmidt, I. Franke, E. Bohnenberger & H. Sandermann, Jr., 'The Cannabinoids: Chemical, Pharmacologic and Therapeutic Aspects', eds. S. Agurell, W. L. Dewey and R. E. Willette, Academic Press, New York – London, 1983, p. 689.
- [4] B. Schmidt, I. Franke, F.-J. Witteler & M. Binder, Helv. Chim. Acta 66, 2564 (1983).
- [5] M. Binder, I. Franke, A. Grothey, S. Koch, F.-J. Witteler & H. Sandermann, Jr., Psychopharmacology, submitted for publication.
- [6] E. Breitmaier & G. Bauer, '13C-NMR-Spektroskopie', Thieme Verlag, Stuttgart, 1977.
- [7] E. Pretsch, Th. Clerc, J. Seibl & W. Simon, 'Tabellen zur Strukturaufklärung organischer Verbindungen', 2nd edn., Springer-Verlag, Berlin – Heidelberg – New York, 1981.